

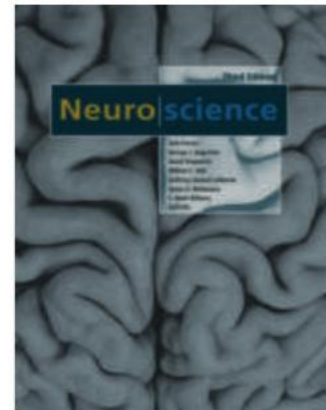
Voltage Gated Ion Channels

The Machines
That Make It
Possible...

Topics I	Topics II
Introduction	Synaptic Transmission
Electrochemical Gradients	Electrophysiology Techniques
Passive Membrane Properties	Basic Circuits (Spinal Cord)
Action Potential	Sensory Systems Overview
Voltage-Gated Ion Channels	Synaptic Plasticity
Ligand-Gated Ion Channels	Recapitulation

Study Material

- NEUROSCIENCE Third Edition
 - Dale Purves
- Chapter 4 pages 69-85



THE COVER

Dorsal view of the human brain.
(Courtesy of S. Mark Williams.)

NEUROSCIENCE: Third Edition

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Aims for this Lecture

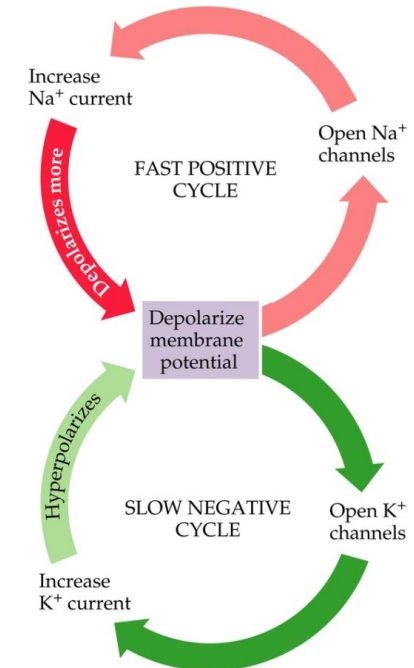
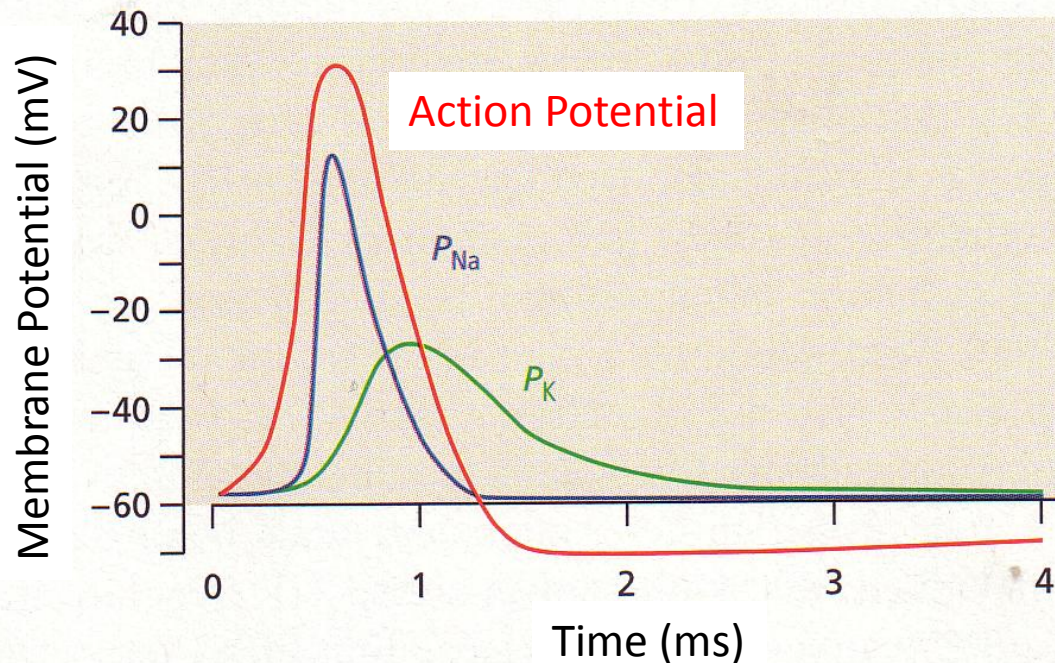
- Get an idea of the molecular structure of voltage-gated ion channels.
- Understand mechanisms for ion selectivity.
- Understand channel opening and closing and its stochastic nature.
- Understand activation and inactivation curves.

Recapitulation L4

- Action potentials are fast stereotypical signals produced by neurons when depolarized above a threshold.
- Na conductances open and let Na flow into the neuron. The neuron further depolarizes. The Na conductances rapidly inactivate.
- K conductances open with a delay and cause an outflow of K ions and a repolarization of the neuron.

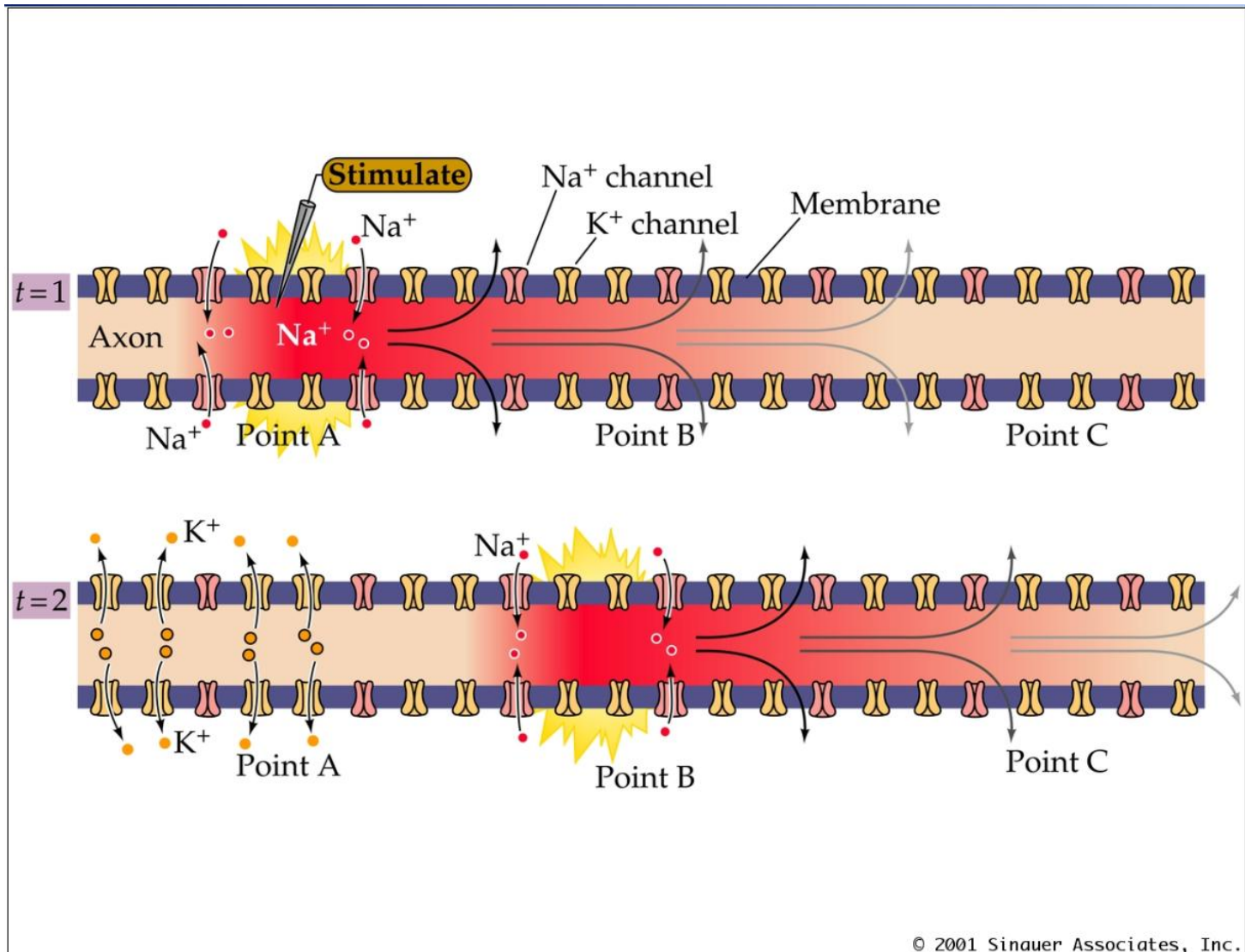
Recapitulation L4

$$E_i = \frac{RT}{F} \ln \left(\frac{P_K [K^+]_a + P_{Na} [Na^+]_a}{P_K [K^+]_i + P_{Na} [Na^+]_i} \right)$$

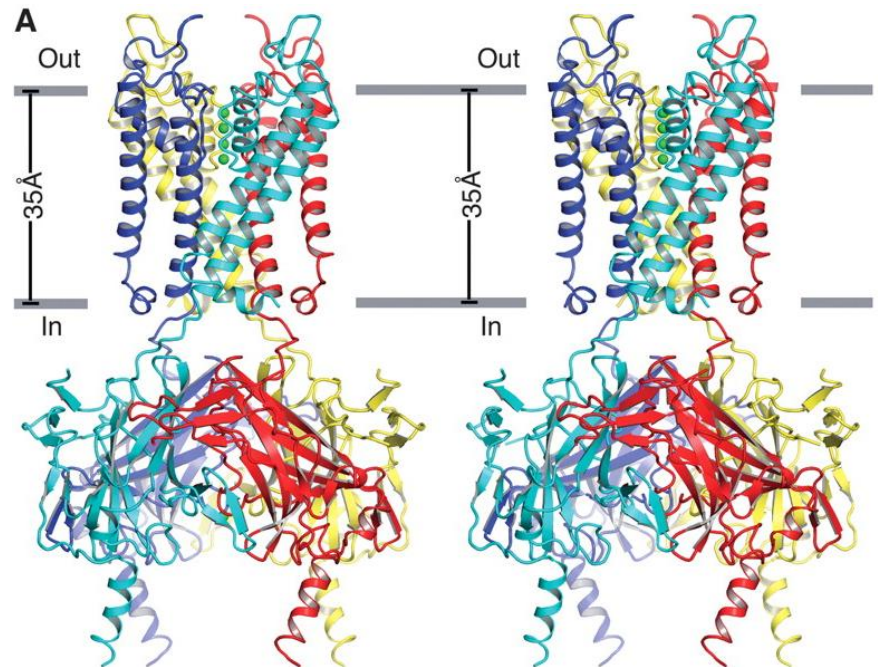
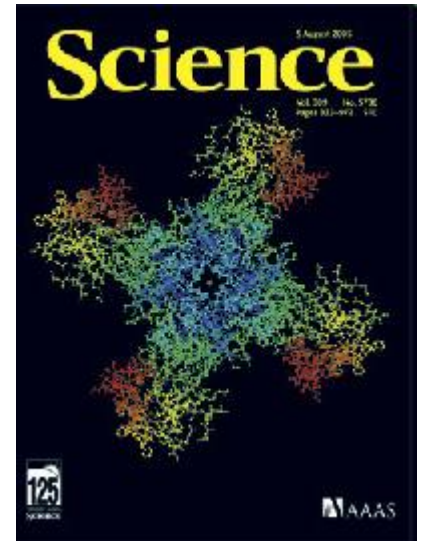
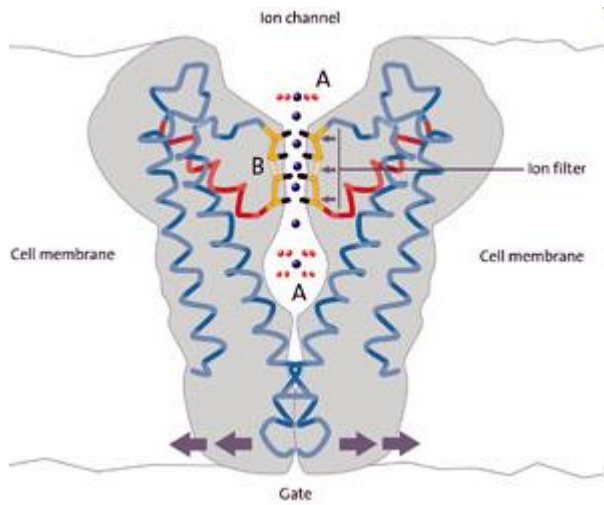


- Positive feedback of voltage dependent Na-conductance
- Negative feedback of voltage dependent K-conductance
- Explosive depolarization and subsequent repolarization..

Recapitulation L4

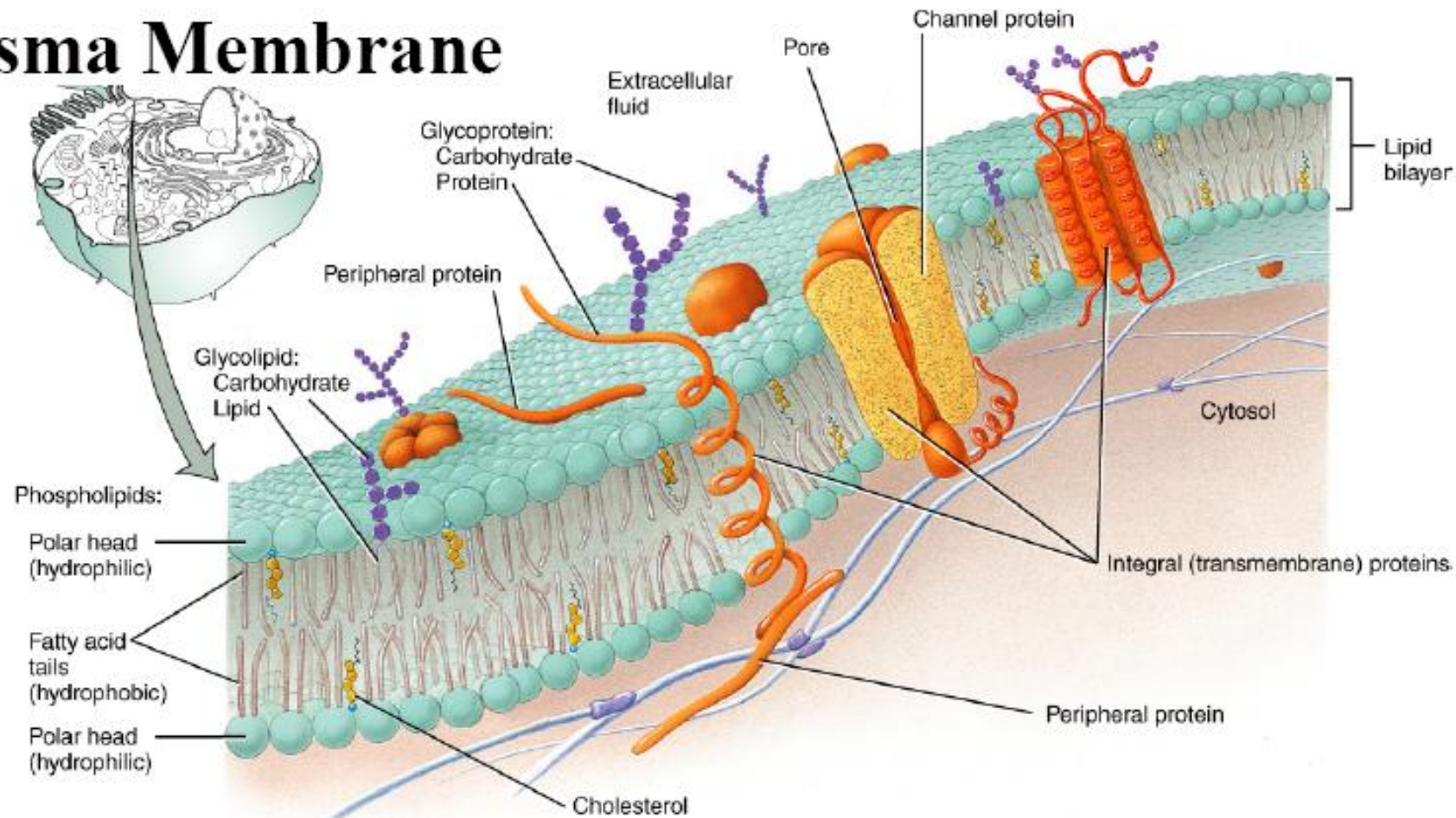


Ion Channels

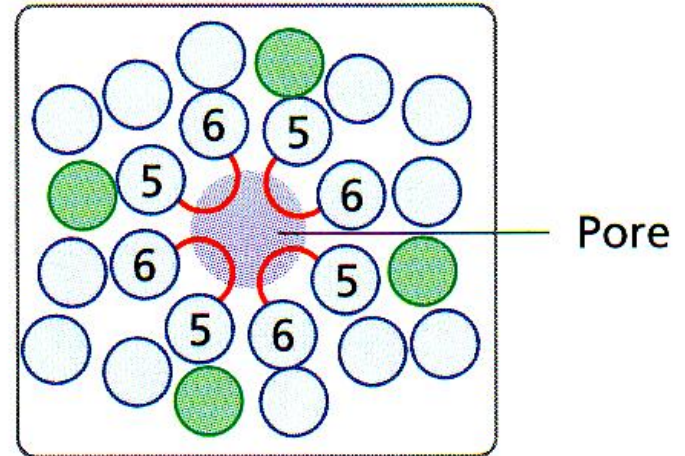
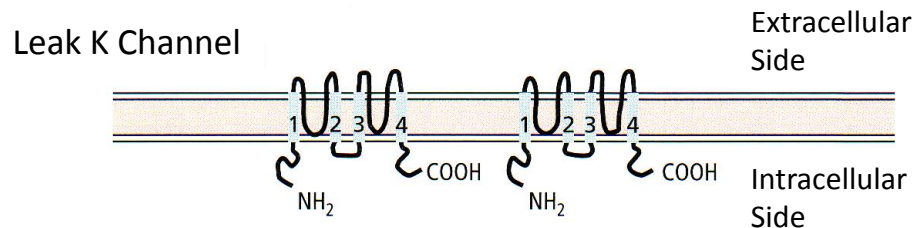
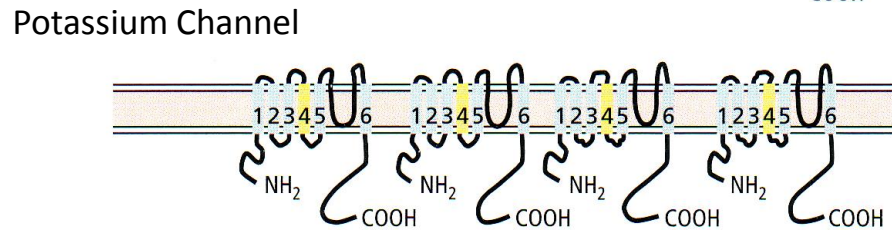
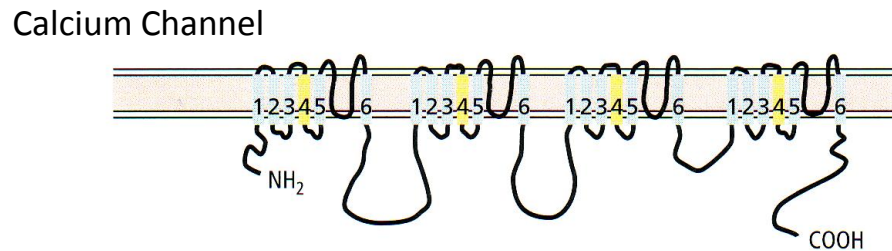
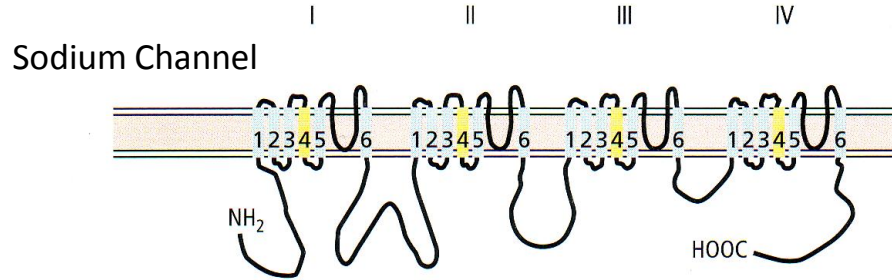


The Cell Membrane

Plasma Membrane

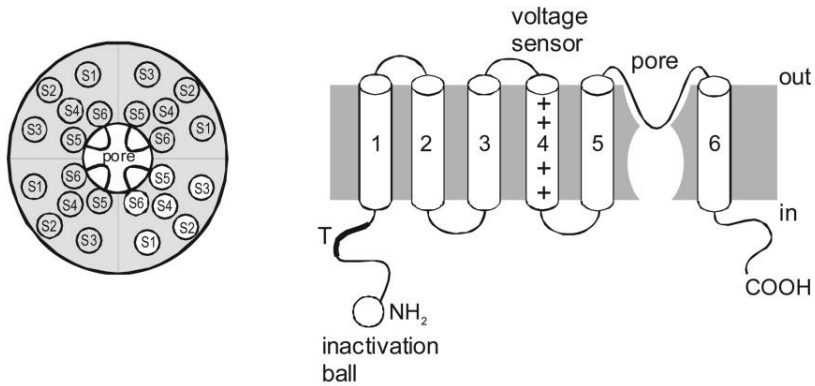


Ion Channels - Structures

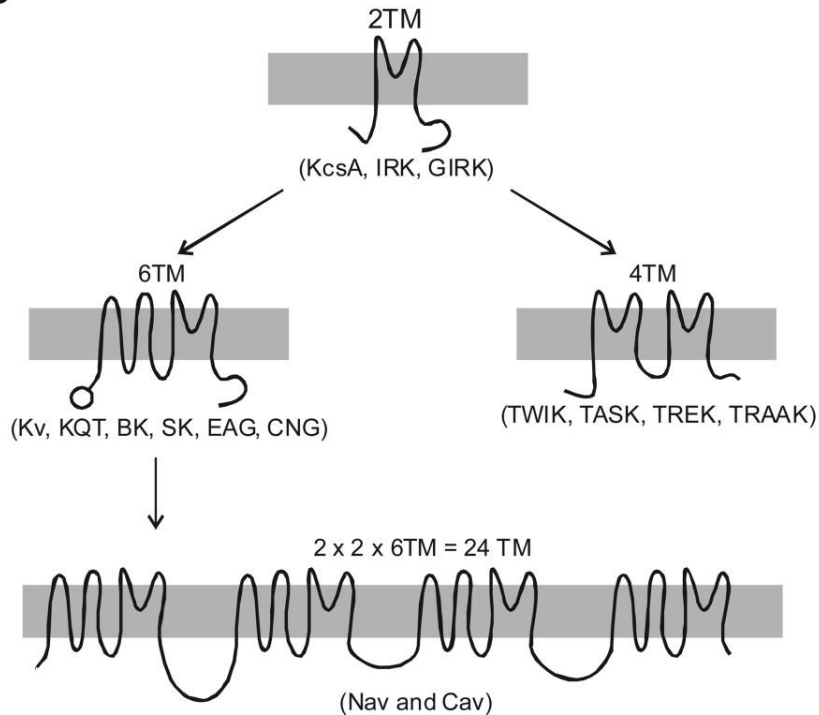


Ion Channel Evolution

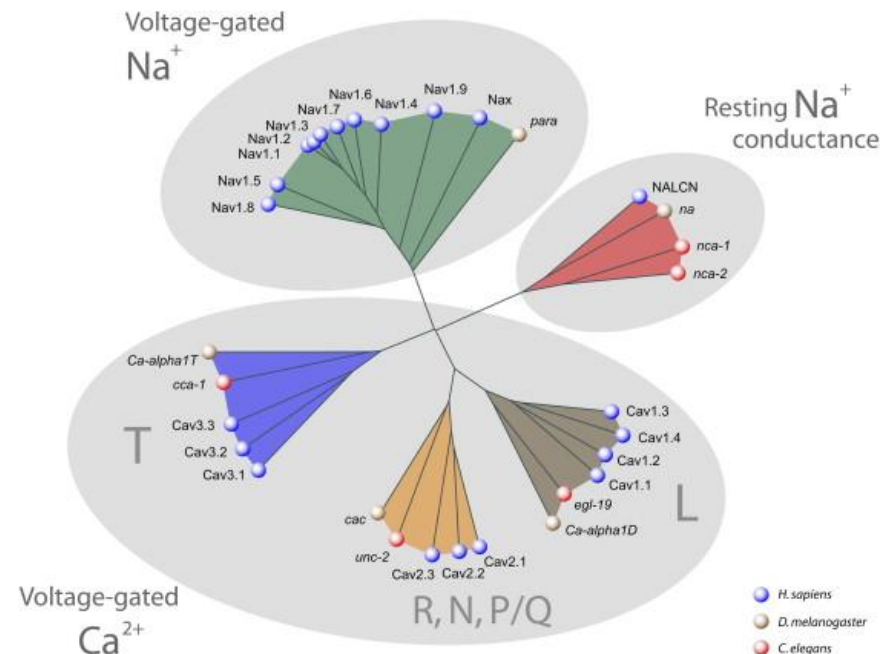
A



B



Four-Domain Channels



Potassium Channels

- The most diverse and most ancient class of ion channels.
- All sequenced genomes contain at least one, most contain many genes for potassium channels.
- Voltage-gated potassium channels are members of this class of channels

K Channels

TABLE 1
K_v channel families

Gene names shown are those assigned by the IUPHAR (Catterall et al., 2002) and HGNC (<http://www.gene.ucl.ac.uk>) in addition to some other commonly used names.

IUPHAR	HGNC	Other
<i>K_v1.1</i>	<i>KCNA1</i>	<i>Shaker</i> -related family
<i>K_v1.2</i>	<i>KCNA2</i>	
<i>K_v1.3</i>	<i>KCNA3</i>	
<i>K_v1.4</i>	<i>KCNA4</i>	
<i>K_v1.5</i>	<i>KCNA5</i>	
<i>K_v1.6</i>	<i>KCNA6</i>	<i>Shab</i> -related family
<i>K_v1.7</i>	<i>KCNA7</i>	
<i>K_v1.8</i>	<i>KCNA10</i>	
<i>K_v2.1</i>	<i>KCNB1</i>	
<i>K_v2.2</i>	<i>KCNB2</i>	
<i>K_v3.1</i>	<i>KCNC1</i>	<i>Shaw</i> -related family
<i>K_v3.2</i>	<i>KCNC2</i>	
<i>K_v3.3</i>	<i>KCNC3</i>	
<i>K_v3.4</i>	<i>KCNC4</i>	
<i>K_v4.1</i>	<i>KCND1</i>	<i>Shal</i> -related family
<i>K_v4.2</i>	<i>KCND2</i>	
<i>K_v4.3</i>	<i>KCND3</i>	
<i>K_v5.1</i>	<i>KCNF1</i>	Modifier
<i>K_v6.1</i>	<i>KCNG1</i>	Modifiers
<i>K_v6.2</i>	<i>KCNG2</i>	
<i>K_v6.3</i>	<i>KCNG3</i>	
<i>K_v6.4</i>	<i>KCNG4</i>	
<i>K_v7.1</i>	<i>KCNQ1</i>	<i>KVLQT</i> <i>EQT2</i>
<i>K_v7.2</i>	<i>KCNQ2</i>	
<i>K_v7.3</i>	<i>KCNQ3</i>	Modifiers
<i>K_v7.4</i>	<i>KCNQ4</i>	
<i>K_v7.5</i>	<i>KCNQ5</i>	
<i>K_v8.1</i>	<i>KCNV1</i>	Modifiers
<i>K_v8.2</i>	<i>KCNV2</i>	
<i>K_v9.1</i>	<i>KCNS1</i>	
<i>K_v9.2</i>	<i>KCNS2</i>	
<i>K_v9.3</i>	<i>KCNS3</i>	
<i>K_v10.1</i>	<i>KCNH1</i>	<i>eag1</i>
<i>K_v10.2</i>	<i>KCNH5</i>	<i>eag2</i>
<i>K_v11.1</i>	<i>KCNH2</i>	<i>erg1</i>
<i>K_v11.2</i>	<i>KCNH6</i>	<i>erg2</i>
<i>K_v11.3</i>	<i>KCNH7</i>	<i>erg3</i>
<i>K_v12.1</i>	<i>KCNH8</i>	<i>elk1, elk3</i>
<i>K_v12.2</i>	<i>KCNH3</i>	<i>elk2</i>
<i>K_v12.3</i>	<i>KCNH4</i>	<i>elk1</i>

TABLE 1
K_{Ca} channels

IUPHAR names of the members of the K_{Ca} group of potassium channels are shown, together with their HGNC designations and other commonly used names.

IUPHAR	HGNC	Other
<i>K_{Ca}1.1</i>	<i>KCNMA1</i>	Slo, Slo1, BK
<i>K_{Ca}2.1</i>	<i>KCNN1</i>	SK _{Ca} 1
<i>K_{Ca}2.2</i>	<i>KCNN2</i>	SK _{Ca} 2
<i>K_{Ca}2.3</i>	<i>KCNN3</i>	SK _{Ca} 3
<i>K_{Ca}3.1</i>	<i>KCNN4</i>	IK _{Ca} 1
<i>K_{Ca}4.1</i>	<i>KCNT1</i>	Slack, Slo2.2
<i>K_{Ca}4.2</i>	<i>KCNT2</i>	Slick, Slo2.1
<i>K_{Ca}5.1</i>	<i>KCNU1</i>	Slo3

BK, big-conductance K⁺ channel; SK, small-conductance K⁺ channel; IK, intermediate-conductance K⁺ channel.

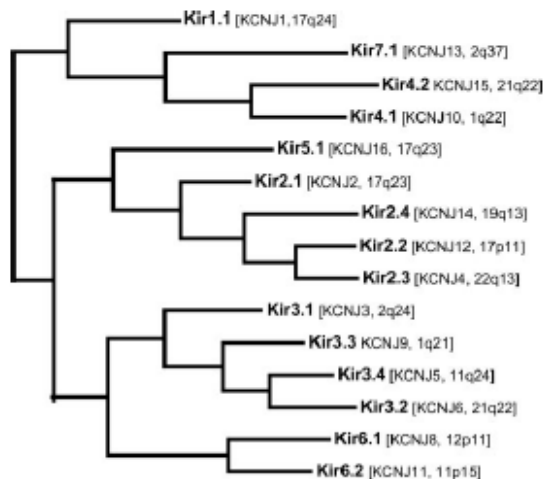


FIG. 1. Phylogenetic tree of K_v channels. Amino acid sequence alignments and phylogenetic analysis for the 15 known members of the human K_v family were generated as described in the legend for Fig. 1 of "LIII. Nomenclature and Molecular Relationships of Voltage-Gated Potassium Channels". No new channels have been added to this topology since it appeared in the earlier edition of this compendium. International Union of Pharmacology and HUGO Gene Nomenclature Committee names of the genes are shown together with their chromosomal localization.

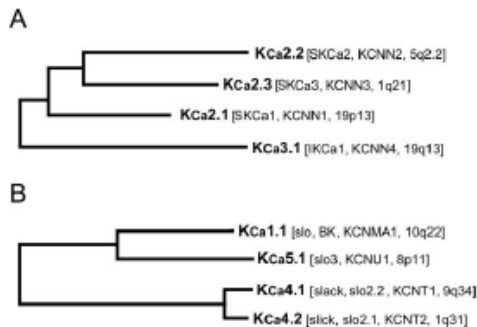
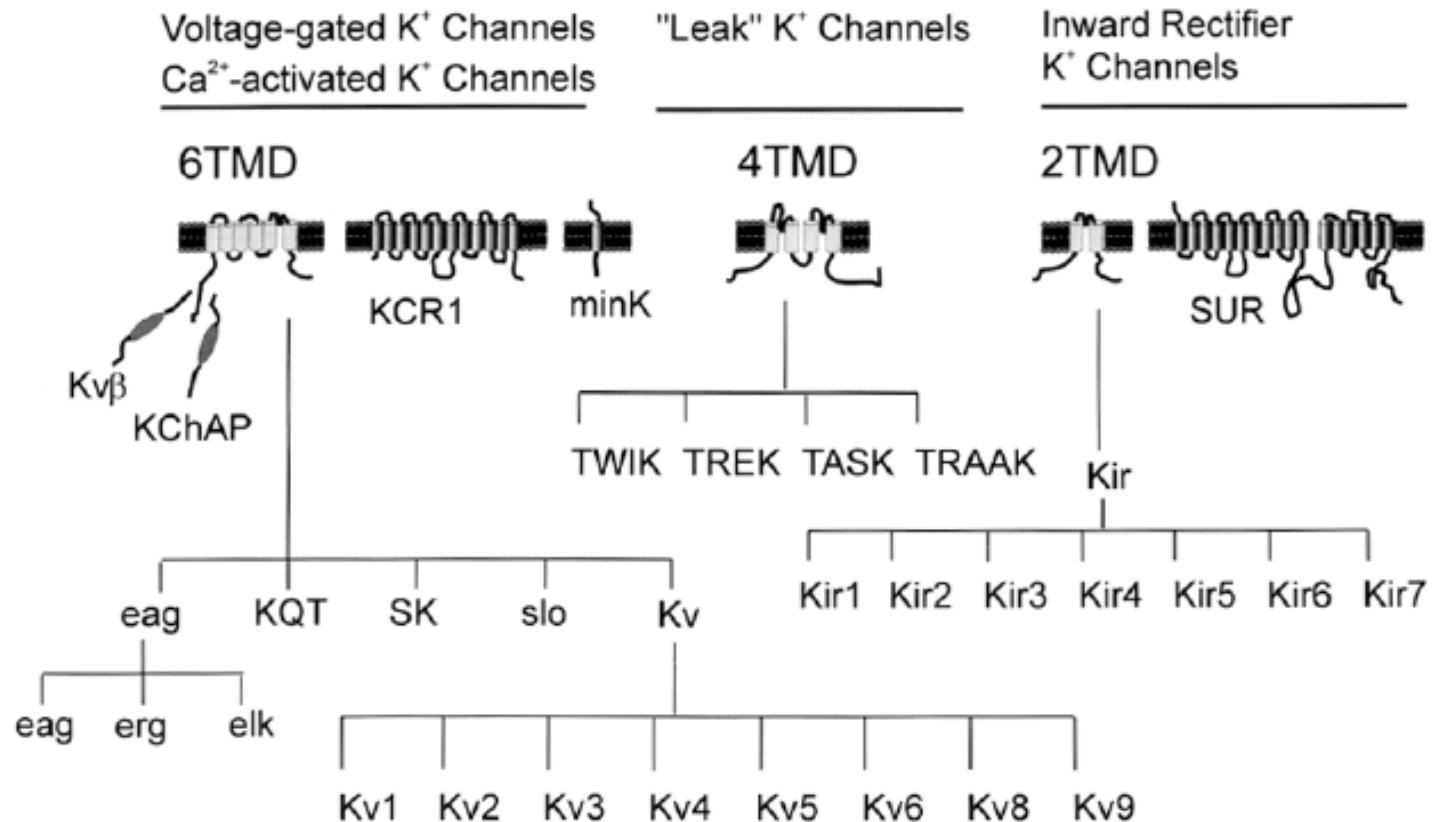


FIG. 1. Phylogenetic tree for K_{Ca} channels. A, K_{Ca}2/3 group. B, K_{Ca}1/4/5 group. Amino acid sequence alignments and phylogenetic analysis for these two groups of four human K_{Ca} channels were generated as described in the legend for Fig. 1 of "International Union of Pharmacology LIII. Nomenclature and Molecular Relationships of Voltage-Gated Potassium Channels". No new channels have been added to these topologies since they appeared in the earlier edition of this compendium. IUPHAR and HGNC names of the genes are shown together with other commonly used names and their chromosomal localization.

Pharmacol Rev. 2005 Dec;57(4):463-72.
Pharmacol Rev. 2005 Dec;57(4):473-508.
Pharmacol Rev. 2005 Dec;57(4):509-26.

Found in all fully sequenced genomes (Archaea, Eubacteria, Eucaryotes).
70 genes in vertebrates code for potassium channels.
Other channels probably evolved from potassium channels.

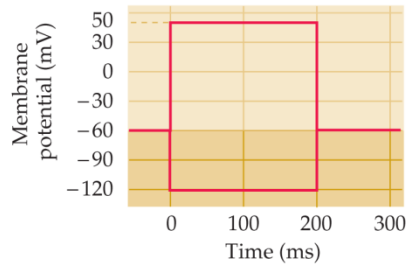
Potassium Channel Classes



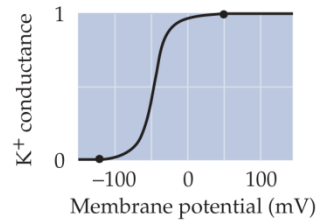
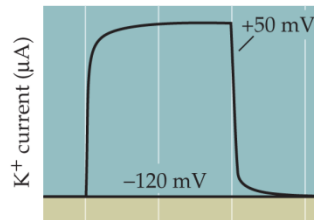
Molecular Diversity of K⁺ Channels

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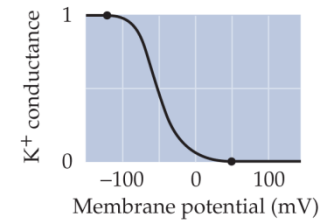
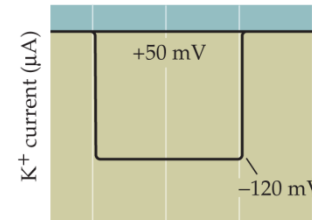
Different K Channels



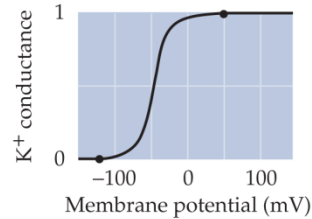
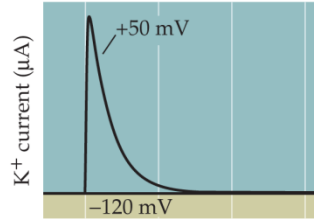
(A) $K_{V2.1}$



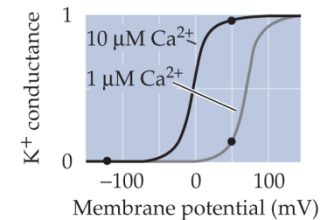
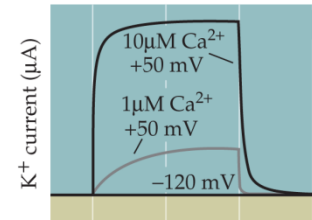
(D) Inward rectifier



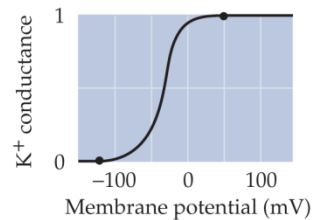
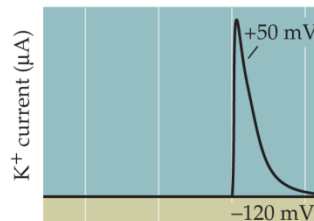
(B) $K_{V4.1}$



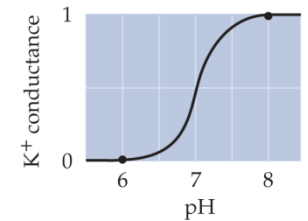
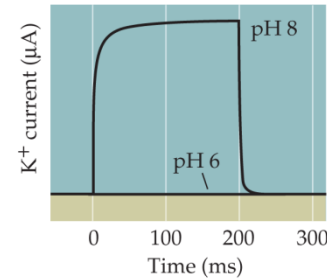
(E) Ca^{2+} -activated



(C) HERG



(F) 2-pore



Voltage-Gated Sodium Channels

- Less diverse – most prominent in excitable tissue that can produce action potentials.
- Four six-transmembrane domain units strung together.
- Prominent inactivation.

Na Channels

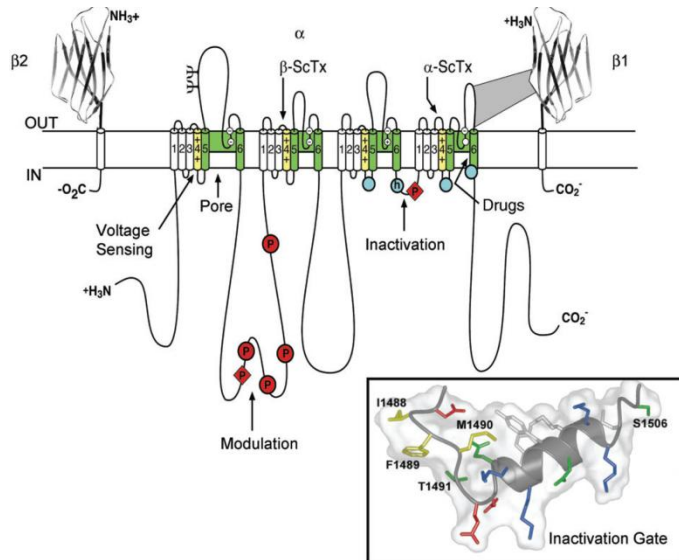


Table 1. Mammalian sodium channel α subunits

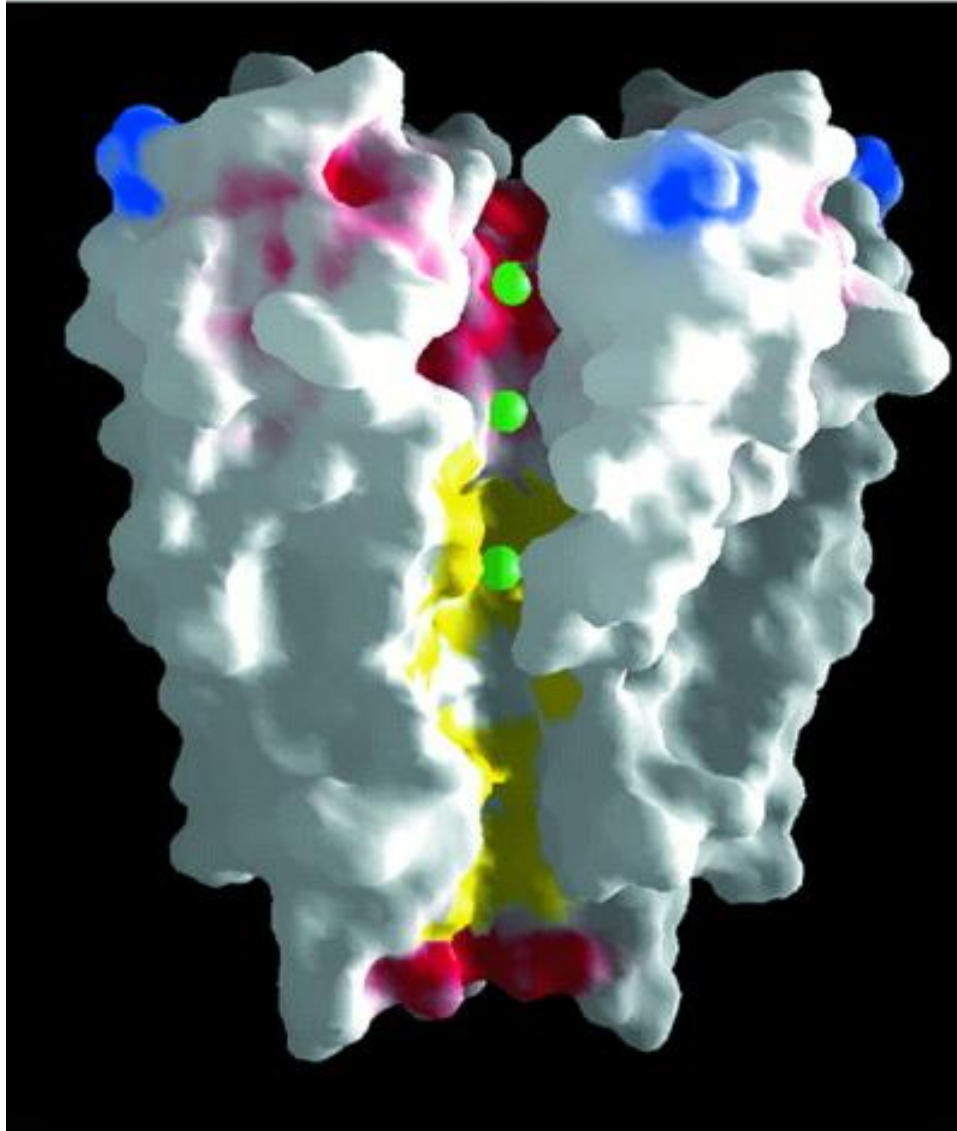
Type	Gene symbol	Chromosomal location	Primary tissues
Na _v 1.1	SCN1A	Mouse 2 Human 2q24	CNS neurons
Na _v 1.2	SCN2A	Mouse 2 Human 2q23–24	CNS neurons
Na _v 1.3	SCN3A	Mouse 2 Human 2q24	CNS neurons
Na _v 1.4	SCN4A	Mouse 11 Human 17q23–25	SkM
Na _v 1.5	SCN5A	Mouse 9 Human 3p21	Uninnervated SkM, heart
Na _v 1.6	SCN8A	Mouse 15 Human 12q13	CNS neurons
Na _v 1.7	SCN9A	Mouse 2 Human 2q24	PNS neurons
Na _v 1.8	SCN10A	Mouse 9 Human 3p22–24	DRG neurons
Na _v 1.9	SCN11A	Mouse 9 Human 3p21–24	DRG neurons
Na _x	SCN7A SCN6A	Mouse 2 Human 2q21–23	uterus, astrocytes, hypothalamus

TOPICAL REVIEW

Voltage-gated sodium channels at 60: structure, function and pathophysiology

William A. Catterall

Ion Selectivity



Blue - positive charges
Red - negative charges
Yellow - hydrophobic

Rough structure of a potassium channel (fourth subunit removed). Negatively charged mouth repels negatively charged ions.

Gruppen	1	2	3
Perioden	s^1	s^2	d^1

Pore Structure

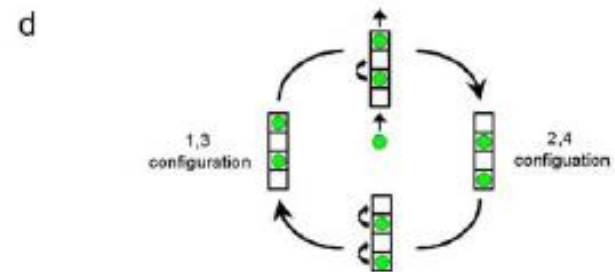
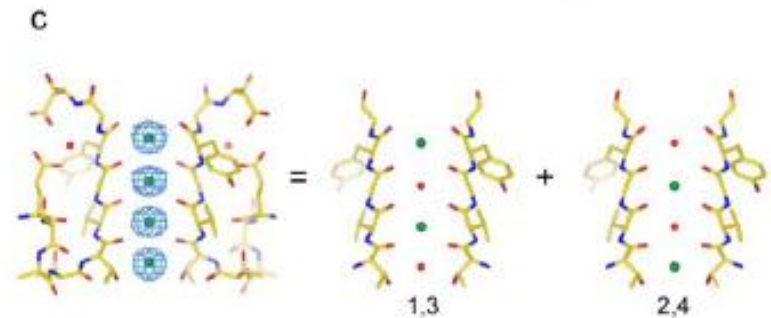
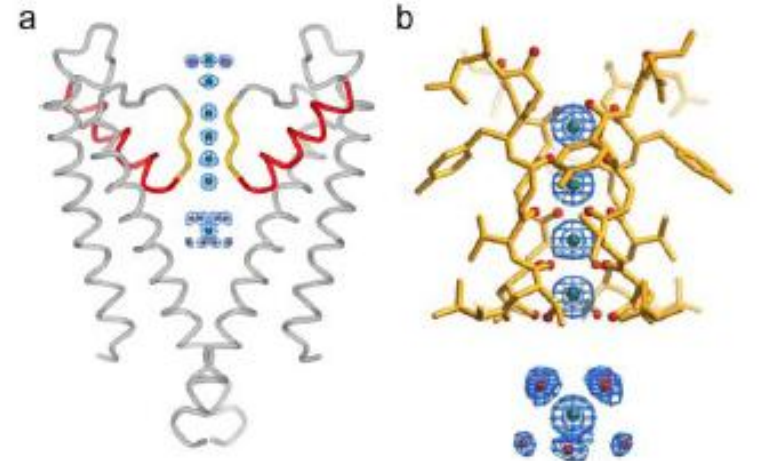
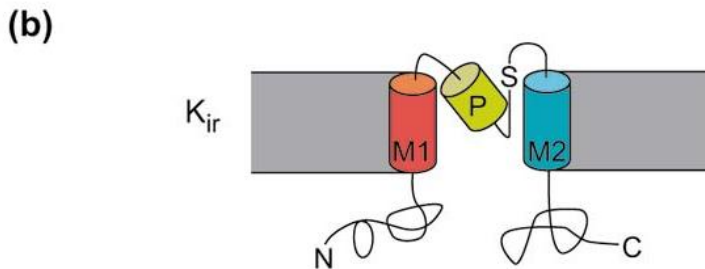
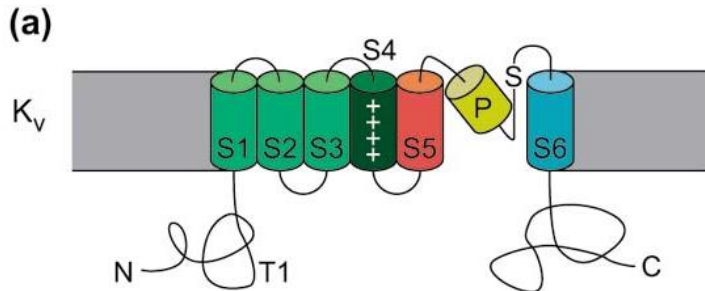
Minireview

Potassium channels

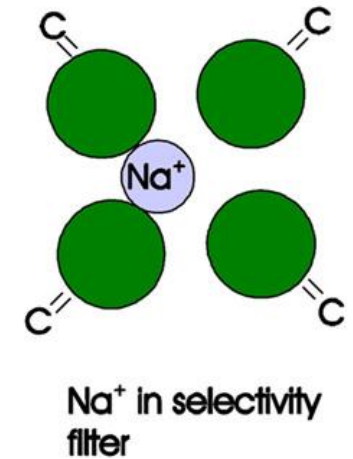
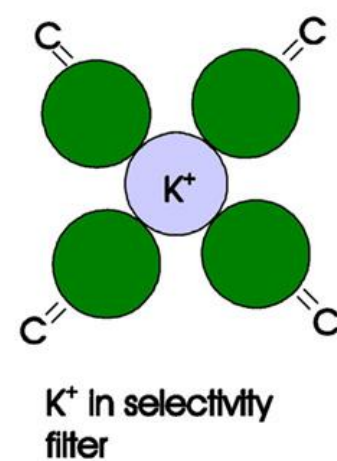
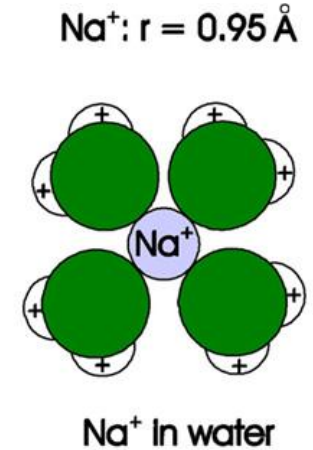
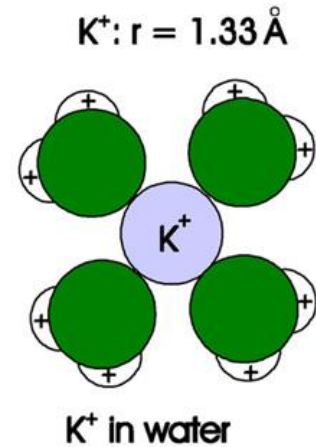
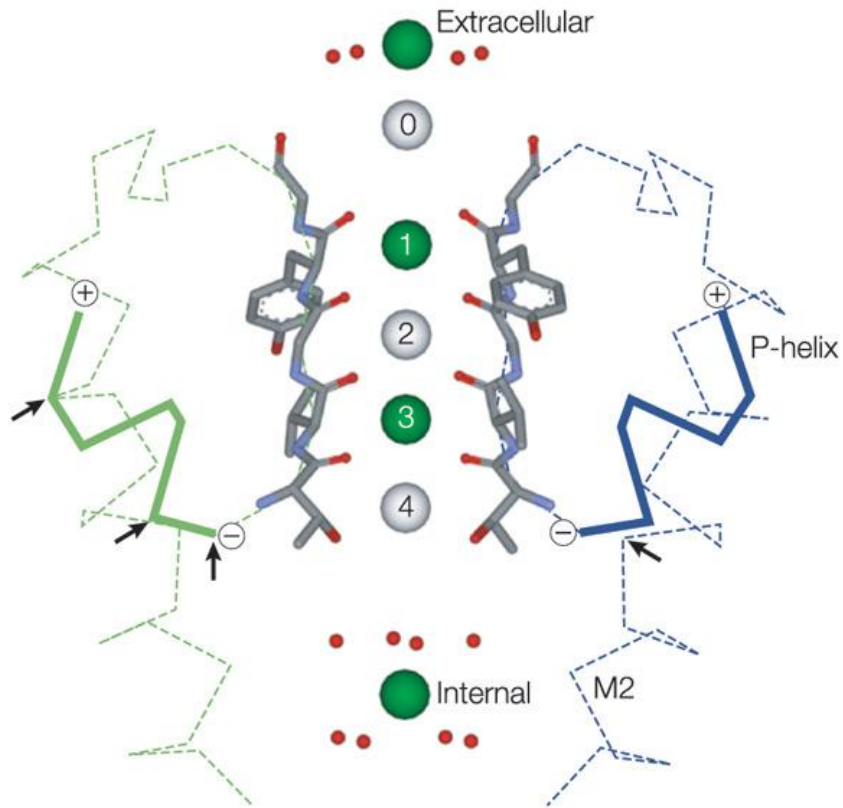
Roderick MacKinnon*

FEBS Letters 555 (2003) 62–6

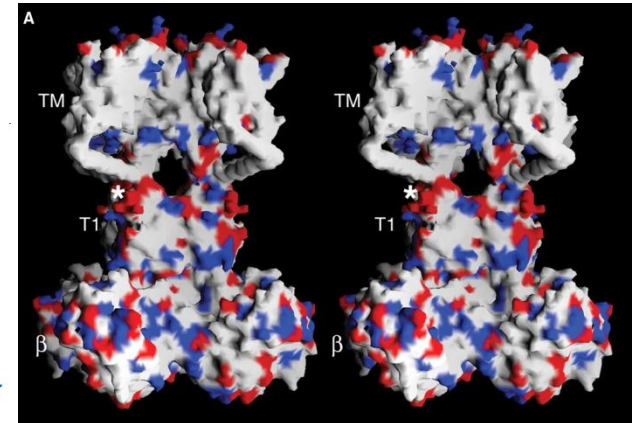
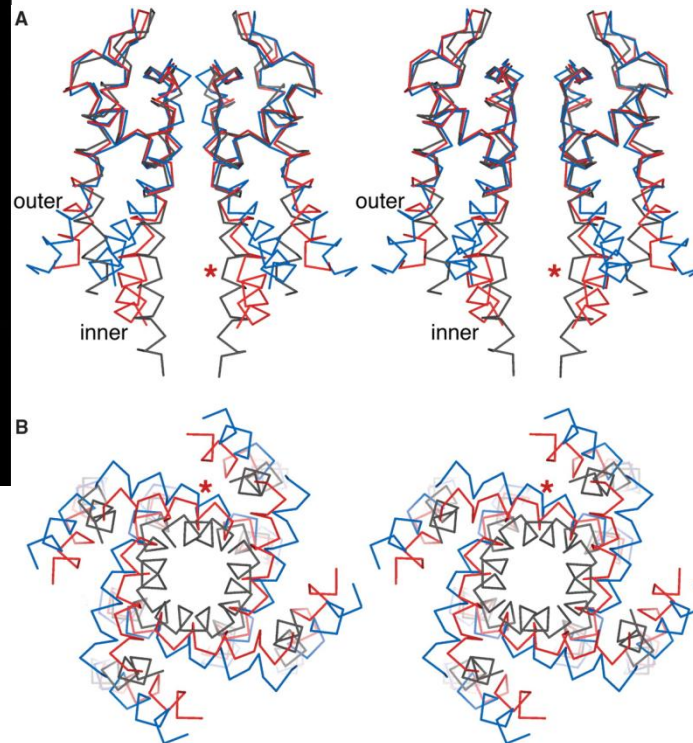
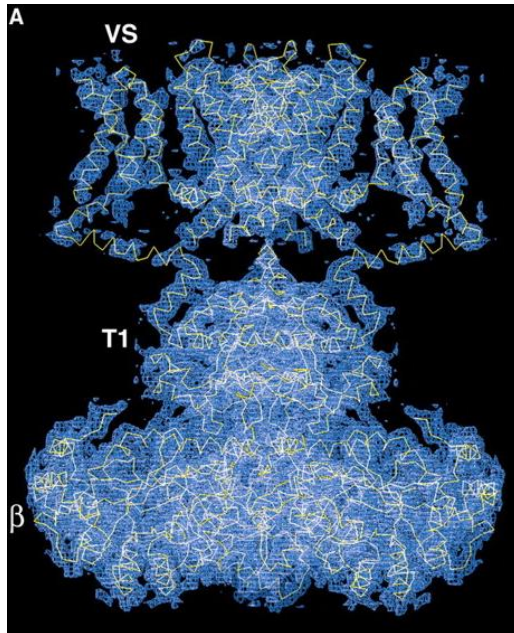
1	1,00 1H		
2	6,94 3Li	9,01 4Be	
3	22,99 11Na	24,31 12Mg	←
4	39,10 19K	40,08 20Ca	44,96 21Sc
5	85,47 37Rb	87,62 38Sr	88,91 39Y
6	132,91 55Cs	137,33 56Ba	138,91 57La*
7	223,02 87Fr	226,83 88Ra	227,03 89Ac#



Selectivity Filter



Mammalian K-Channel



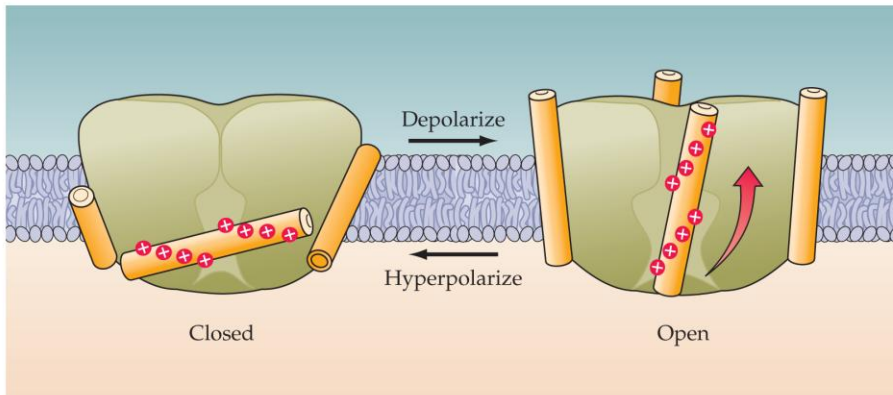
Science 5 August 2005:
Vol. 309, no. 5736, pp. 897 - 903

Crystal Structure of a Mammalian Voltage-Dependent *Shaker* Family K⁺ Channel
Stephen B. Long, Ernest B. Campbell, Roderick MacKinnon*

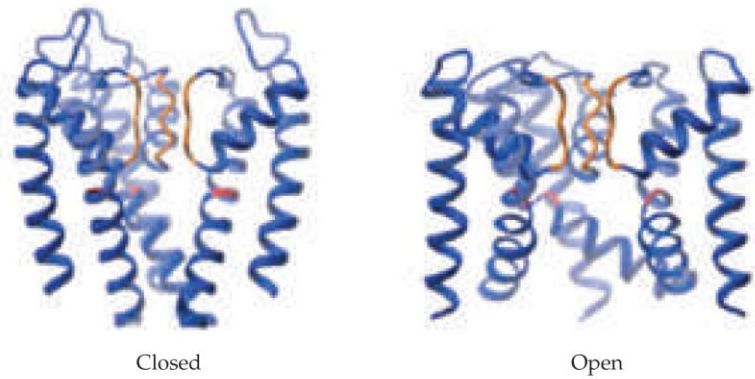
3D if watched cross-eyed

K Channel Gating

(A)



(B)



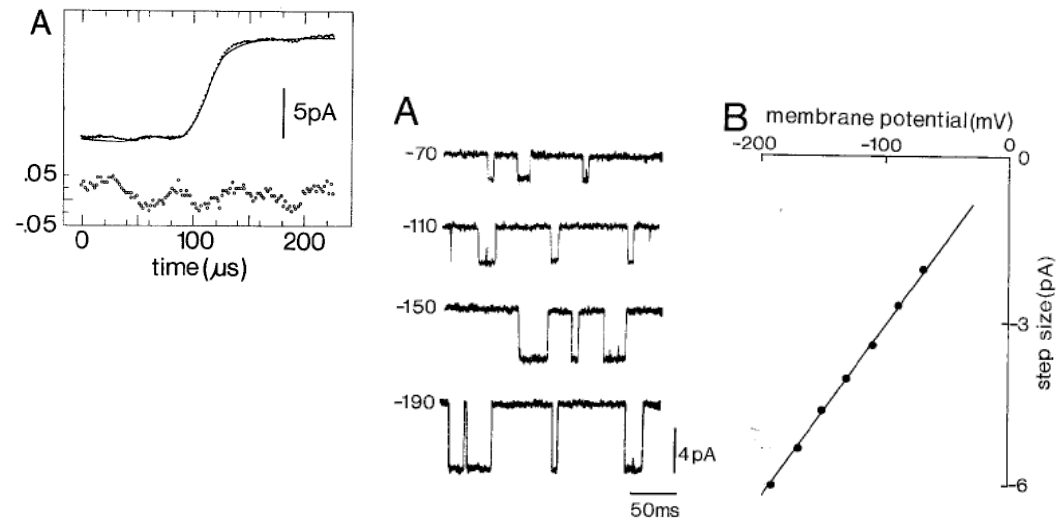
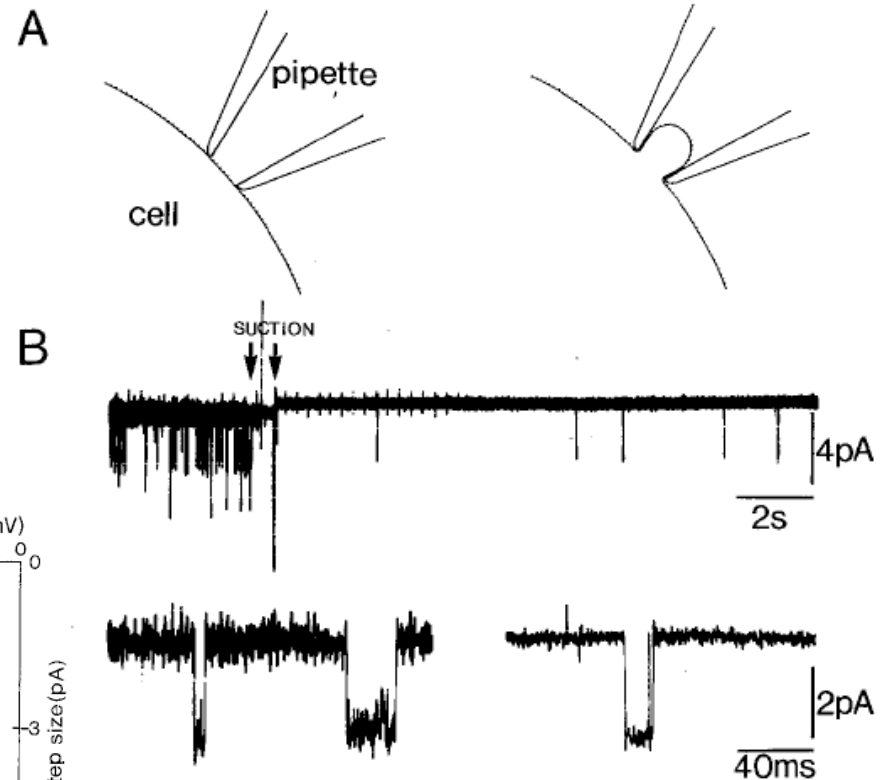
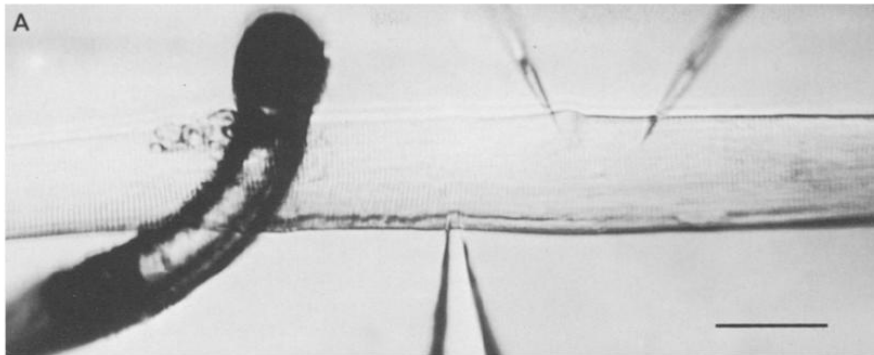
Measuring Individual Channels

Pflügers Arch (1981) 391:85–100

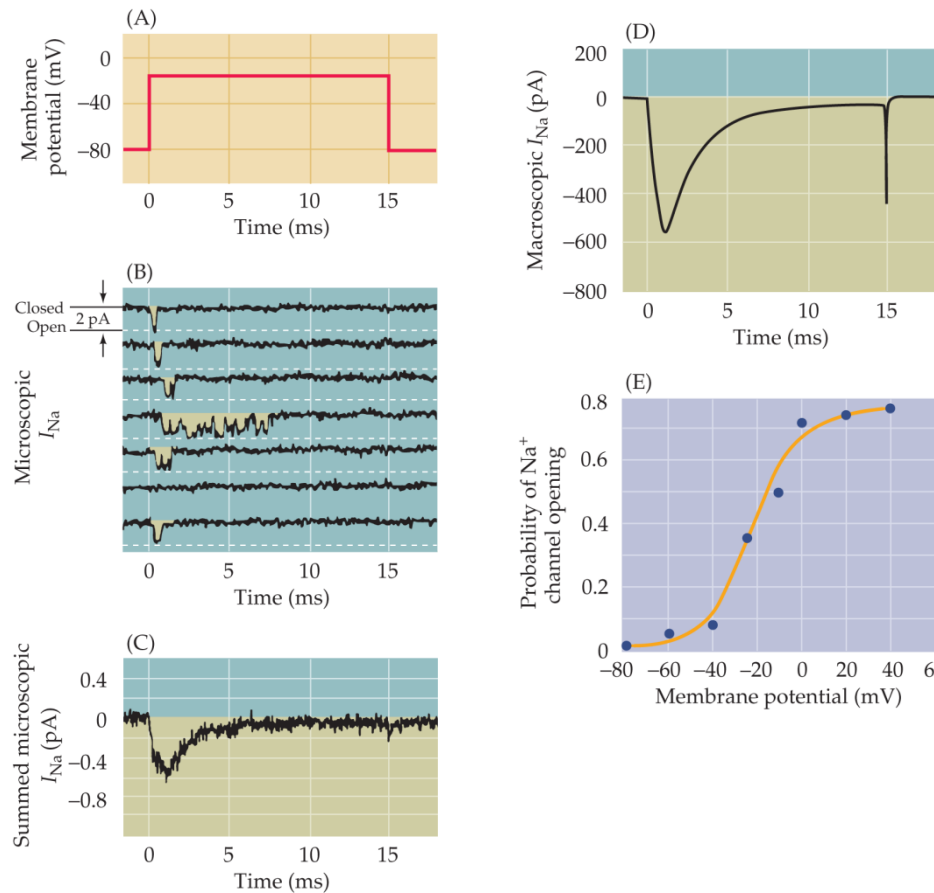
Improved Patch-Clamp Techniques for High-Resolution Current Recording from Cells and Cell-Free Membrane Patches

O. P. Hamill, A. Marty, E. Neher, B. Sakmann, and F. J. Sigworth

Max-Planck-Institut für biophysikalische Chemie, Postfach 968, Am Fassberg, D-3400 Göttingen, Federal Republic of Germany

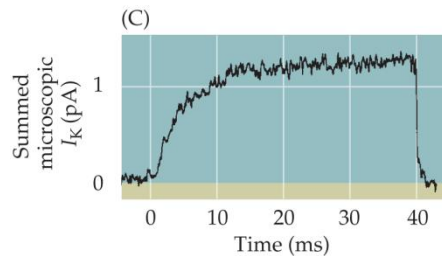
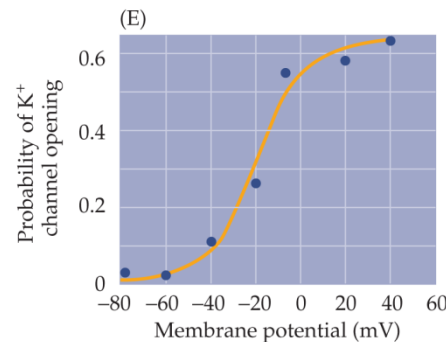
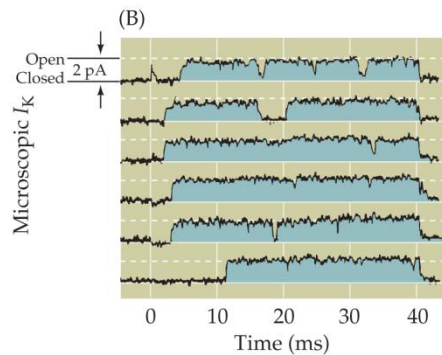
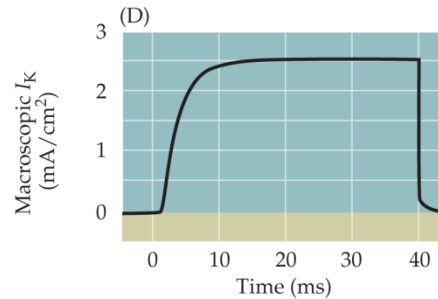
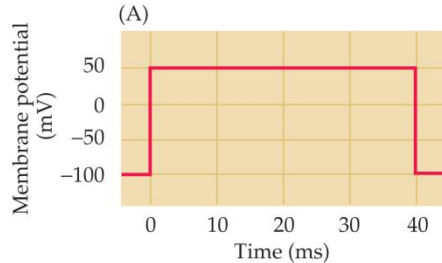


Na Channel Behavior



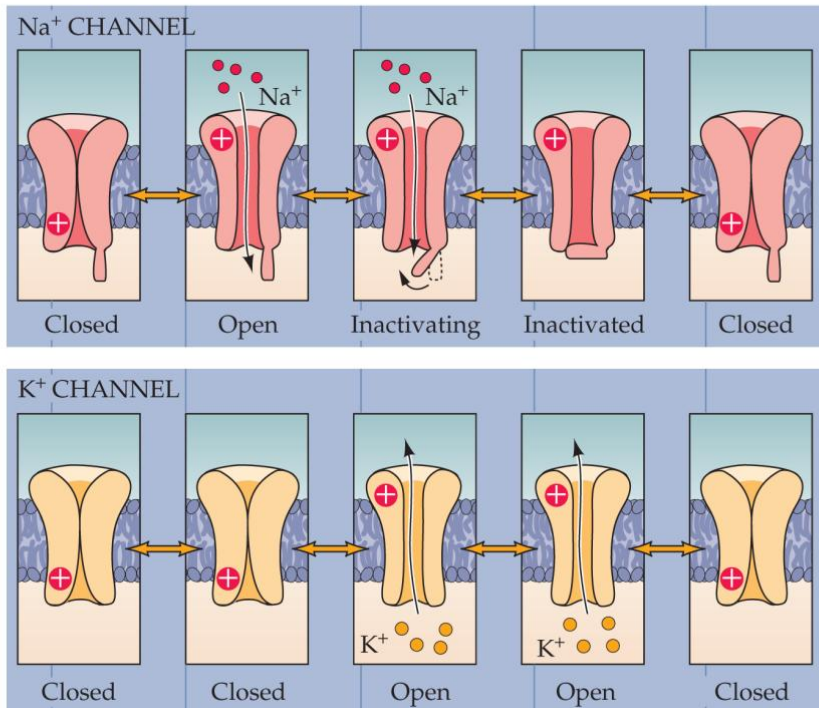
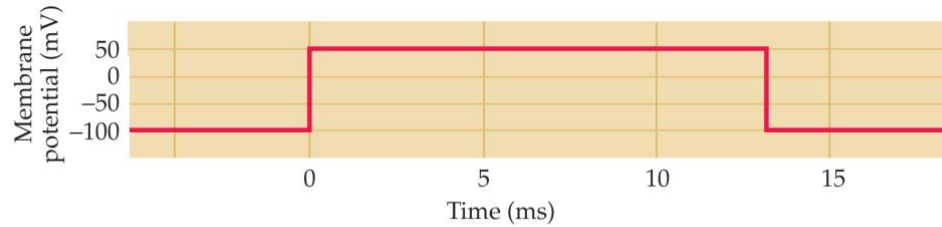
Individual channels behave stochastically – only the probability of their opening can be predicted. Because many channels are typically involved the macroscopic current can still be predicted with high accuracy

K Channel Behavior

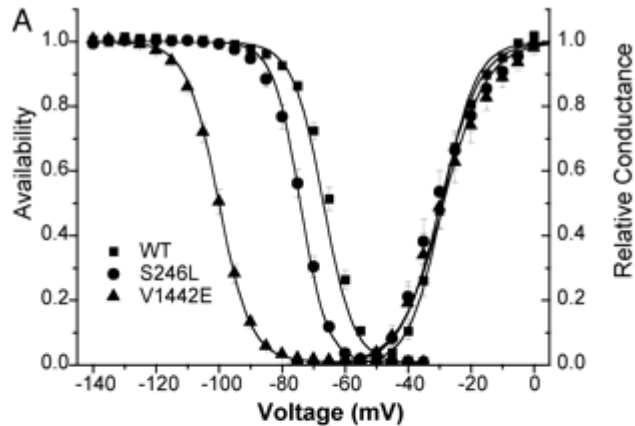


Individual channels behave stochastically – only the probability of their opening can be predicted. Because many channels are typically involved the macroscopic current can still be predicted with high accuracy

AP and Channel States



Activation and Inactivation Curves



Myasthenic syndrome caused by mutation of the *SCN4A* sodium channel

PNAS June 10, 2003 vol. 100

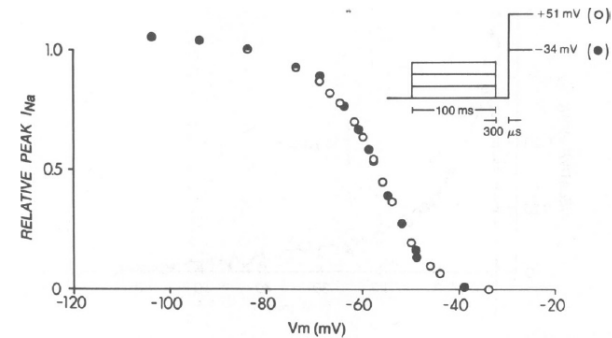


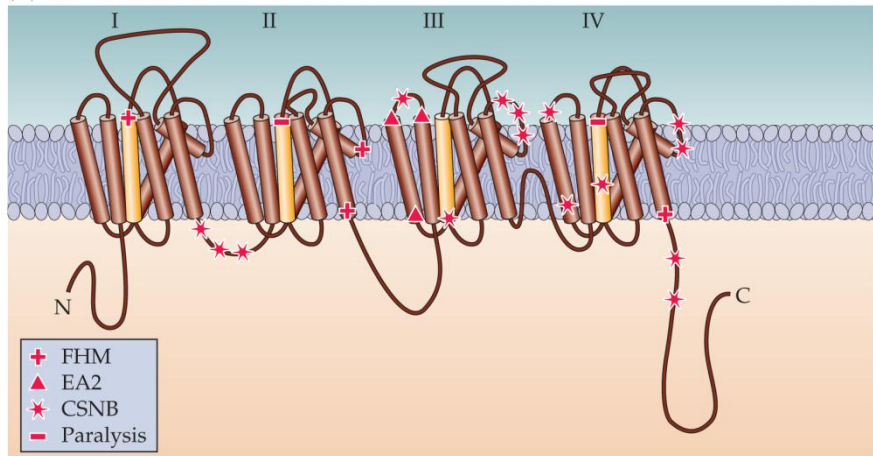
FIGURE 9 Absence of shift in steady-state activation curve with test-pulse height. Steady-state inactivation curves were obtained by giving a 100-ms prepulse to a variable potential, returning to the holding potential for 300 μ s, and stepping to the test-pulse potential. The peak currents during the test pulses are plotted as a function of the prepulse potential, normalized to the value for a step from -84 mV, the holding potential. The data points for -94 and -104 mV for the -34-mV test pulse show that there was virtually no inactivation at -84 mV; peak current was about 5% increased by the 100-ms prepulse to -104 mV, and much of this may have been due to removal of slow inactivation. Peak conductance at -34 mV was 0.19 of that at +51 mV. $\frac{1}{2}$ Na, $\frac{1}{2}$ K, 1 mM 4-AP, 3.3–3.5°C.

**SODIUM CHANNEL INACTIVATION IN THE
CRAYFISH GIANT AXON
MUST CHANNELS OPEN BEFORE INACTIVATING?**

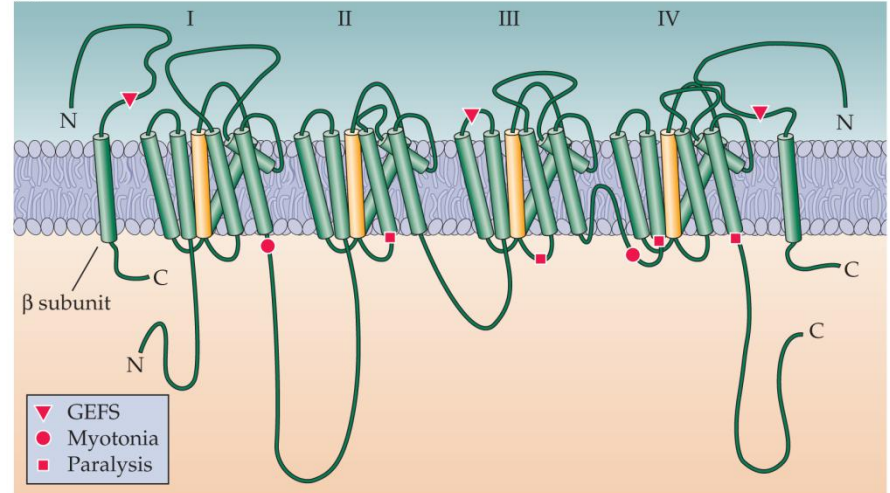
BIOPHYS. J. © Biophysical Society • 0006-3495/09/81/595/20 \$1.00
Volume 35 September 1981 595-614

Channelopathy

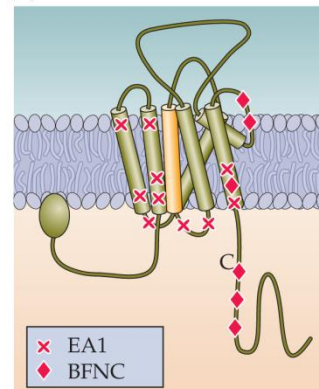
(A) Ca^{2+} CHANNEL



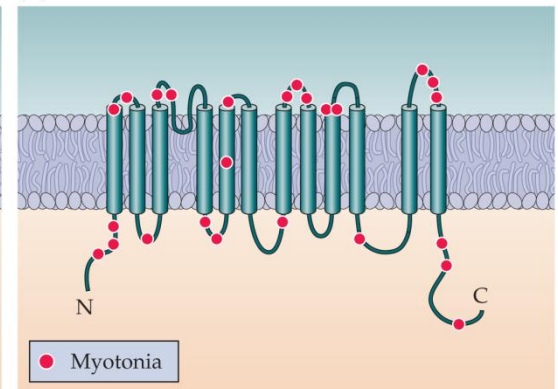
(B) Na^{+} CHANNEL



(C) K^{+} CHANNEL



(D) Cl^{-} CHANNEL



Familial Hemiplegic Migraine
 Episodic Ataxia Type 2
 Congenital Stationary Night Blindness

Generalized Epilepsy with Febrile Seizures
 Benign Familial Neonatal Convulsion

What Do You Want To Know....

- ... about a channel?
- How is it opened (activation curve)?
- How is it closed (de-activation, inactivation curve)?
- Which ion type(s) go through?
- What are its kinetics?
- What is its single channel conductance?

These parameters may vary depending on subunit assembly, phosphorylation, ion-, cAMP-, ATP-binding,